

# FAVORGEN® FavorPrep™ 96-well Gel/ PCR Clean-Up DNA Kit

(For Research Use Only)

Kit Contents/ Cat. No.:	FAPKE96001 1 plate	FAPKE96002 2 plates	FAPKE96004 4 plates
Binding Buffer D1	60 ml	135 ml	135 ml x 2
Wash Buffer (concentrated)	35 ml <sup>†</sup>	35 ml $\times 2^{\dagger}$	$35 \text{ ml x } 4^{\dagger}$
Elution Buffer	20 ml	50 ml	50 ml x 2
Filter Plate (96-Well DNA binding plate	) 1 plate	2 plates	4 plates
96-Well 2 ml Plate	3 plate	6 plates	12 plates
96-Well PCR plate	1 plate	2 plates	4 plates
Adhesive Film	2 pcs	4 pcs	8 pcs

<sup>+</sup>Add 140 ml ethanol (96~100%) to each Wash Buffer when first open.

# **Quality Control**

The quality of 96-Well Gel/ PCR Clean-Up DNA Kit is tested on a lot-to-lot basis. The purified DNA is checked by agarose gel analysis and quantified with spectrophotometer.

# **Specification:**

Sample Size: up to 200 mg agarose gel slice

up to 100 µl PCR or other enzymatic reaction mixture

Binding Capacity: up to 20 µg/ well DNA Size range: 70 bp ~12Kb Minimum elution volume: 50 µl

Operation: centrifuge/vacuum manifold

Handling Time: about 40 minutes for gel DNA extraction

about 30 minutes for PCR clean up

Recovery: 70~85 % for gel DNA extraction

90~95 % PCR clean up

Downstream application: Fluorescent or radioactive sequencing, Restriction digestion,

Library screening, Ligation, Labeling, Transformation

# **Important Notes:**

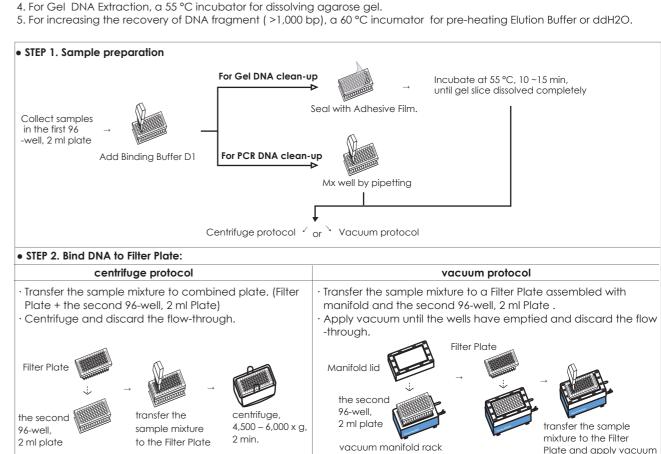
- 1. Buffers provided in this system contain irritants. Wear gloves and lab coat when handling these buffers.
- 2. The maxium sample size is described on specification, do not use the sample more than the limitation.
- 3. When excising the agarose gel, remove the extra gel to minimize the size of the gel.
- 4. Add ethanol (96~100 %) to Wash Buffer when first open.
- 5. Prepare bath to 55 °C before the operation for Gel DNA Extraction Protocol.
- 6. Preheat the Elution Buffer to 60 °C for DNA elution to increase the recovery of DNA fragment >1,000 bp.



# **Brief procedure:**

#### Material to be provided by user for a 96-well preparation

- 1. Centrifuge equiment with a swing -bucket rotor and adaptor for 96-well plate, capable of at least 5,600 ~ 6,000 X g.
- 2. Vacuun manifold for 96-well plate and Vaccum source for 10 inches Hg.
- 3.96~100 % ethanol for preparation of the Wash Buffer when first use.
- 4. For Gel DNA Extraction, a 55 °C incubator for dissolving agarose gel.



# • STEP 3. Wash Filter Plate twice with Wash Buffer

1st Washing: Add Wash Buffer and centrifuge 5,600 -6,000 x g, 2 min.

2nd Washing: Add Wash Buffer and centrifuge 5,600 6,000 x g, 15 min.





1st Washing: Add Wash Buffer and apply vacuumat 10 inches Ha for until the wells have emptied.

at 10 inches Hg.

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2nd Washing: add Wash Buffer and apply vacuumat 10 inches Hg for 10 min.

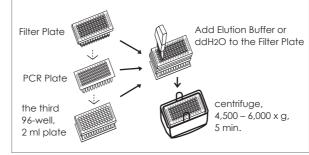


#### STEP 4 . Dry the membranes of Filter Plate:

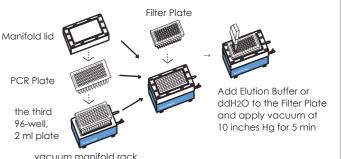
- · Stand the Filter plate on a clean paper towel at room temperature 10 min.
- Gently tap the tips of the Filter Plate on paper towel to remove residual liquid
- Apply vacuum at 10 inches Hg for an addition 10 min.

## • STEP 5. DNA Elution:

- Add Elution Buffer or ddH2O to the Filter Plate. Stand for 3 min.
- Centrifuge to elute DNA.



Add Elution Buffer or ddH2O to the Filter Plate. Stand for 3 min. Apply vacuum to elute DNA.



# Protocol: (centrifugation processing)

- using centrifuging force to handle DNA binding step and washing steps.

# Material to be provided by user for a 96-well preparation

- 1. Centrifuge equiment with a swing -bucket rotor and adaptor for 96-well plate, capable of at least 5,600 ~ 6.000 X a.
- 2. Preheat required Elution Buffer or ddH2O (40 ~100 µl per well) at 60 °C. (For Step 4 DNA elution) -- For increasing the recovery of DNA fragment (>1,000 bp).

## Please Read Important Notes Before Starting The Following Steps.

## **STEP 1: Sample Preparation**

#### For Gel DNA Clean-Up

- Transfer up to 200 mg of agarose gel (containing relevant DNA fragment) to each well of a 96-Well 2 ml Plate (provided, the first 96-well, 2 ml plate).
- Add 500 µl of Binding Buffer D1 to each well, sealing with a adhesive film. Incubate at 55 °C for 10~15 minutes until the gel slice dissolved completely. During the incubation, briefly shake the incubated plate for every 5 minutes to make the sample mixture mix well with Binding Buffer D1.

## For PCR DNA Clean-Up

- Transfer up to 100 µl of PCR or enzymatic product to each well of a clean 96-Well 2 ml Plate. (provided, the first 96-well, 2 ml plate).
- · Add 5 volume of Binding Buffer D1 to each well. Mix well by pipetting.
- For example, Add 250 µl of Binding Buffer D1 to 50 µl of PCR or enzymatic product.

# STEP 2: DNA Binding

- Place a Filter Plate (96-Well DNA Binding Plate) on top of a clean 96-well plate (provided, the second 96-Well 2 ml Plate).
- · Transfer the sample mixture to the Filter Plate.
- Place the combined plate (Filter Plate combined with the second 96-Well, 2 ml plate) in a rotor bucket and centrifuge at 4,500 6,000 x g for 2 min.
- $\cdot$  Discard the flow-through and return the Filter Plate back to the second 96-Well 2 ml Plate.

# STEP 3: Wash Filter Plate Twice (W1 Buffer and Wash Buffer)

#### 1st Washing

- $\cdot$  Add 650  $\bar{\mu}l$  of Wash Buffer (ethanol added) to each well of the Filter Plate.
- Place the combined plate (Filter Plate combined with the second 96-Well 2 ml plate) in a rotor bucket and centrifuge at 4,500 6,000 xg for 2 min.
- · Discard the flow-through and return the Filter Plate back to the second 96-Well 2 ml Plate.

# 2nd Washig

- $\cdot$  Add 650  $\mu$ I of Wash Buffer to each well of the Filter Plate.
- · Place the combined plate in a rotor bucket and centrifuge at 4,500 6,000 xg for 15 min.
- $\cdot$  Discard the second 96-Well 2 ml plate and the flow-through.

# STEP 4: Dry the membranes of Filter Plate

· Stand the Filter plate on a clean paper towel at room temperature 10 min.

# STEP 5: DNA Elution

- $\cdot$  Place a 96-well PCR Plate (provided) on top of a clean 96-Well 2 ml Plate. (provided, the third 96-well, 2 ml plate). and place the Filter Plate on the 96-Well PCR plate.
- (top: Filter Plate, middle: 96-well PCR Plate, bottom: 96-Well 2 ml plate)
- $\cdot$  Add 50  $\sim$  100  $\mu$ l of Elution Buffer or ddH<sub>2</sub>O (pH8.0-8.5) to the membrane center of the Filter Plate. Stand for 3 min until Elution Buffer or ddH<sub>2</sub>O has been absorbed by the membrane completely.
- Place the combined plate in a rotor bucket and centrifuge at 4,500 6,000 x g for 5 min to elute DNA to the 96-well PCR plate.

# Protocol: (Vacuum processing)

- using vacuum force to handle DNA binding step and washing steps.

# Material to be provided by user for a 96-well preparation

- 1. Vacuun manifold for 96-well plate and a vaccum source for 10 inched Hg.
- 2. Preheat required Elution Buffer or ddH2O (40 ~100 µl per well) at 60 °C. (For Step 4 DNA elution)
  - -- For increasing the recovery of DNA fragment (>1,000 bp).

## Please Read Important Notes Before Starting The Following Steps.

#### **STEP 1: Sample Preparation**

#### For Gel DNA Clean-Up

- Transfer up to 200 mg of agarose gel (containing relevant DNA fragment) to each well of a 96-Well 2 ml Plate (provided, the first 96-well, 2 ml plate).
- Add 500 µl of Binding Buffer D1 to each well, sealing with a adhesive film. Incubate at 55 °C for 10~15 minutes until the gel slice dissolved completely. During the incubation, briefly shake the incubated plate for every 5 minutes to make the sample mixture mix well with Binding Buffer D1.

## For PCR DNA Clean-Up

- Transfer up to 100 µl of PCR or enzymatic product to each well of a clean 96-Well 2 ml Plate. (provided, the first 96-well, 2 ml plate).
- · Add 5 volume of Binding Buffer D1 to each well. Mix well by pipetting.
- For example, Add 250 µl of Binding Buffer D1 to 50 µl of PCR or enzymatic product.

## **STEP 2: DNA Binding**

- · Place a clean 96-well plate (provided, the second 96-Well, 2 ml Plate) on the rack of vacuum manifold and cover the manifold lid.
- · Place a Filter Plate (96-well DNA Binding Plate, provided) on top of the second 96-Well 2 ml plate.
- · Transfer the sample mixture to the Filter Plate.
- · Apply vacuum until the wells have emptied.

# STEP 3: Wash Filter Plate Twice (W1 Buffer and Wash Buffer)

## 1st Washing

- $\cdot$  Add 650  $\mu$ l of Wash Buffer (ethanol added) to each well of the Filter Plate.
- · Apply vacuum at 10 inches Ha until the wells have emptied.

# 2nd Washing

- $\cdot$  Add 650  $\mu$ l of Wash Buffer to each well of the Filter Plate.
- · Apply vacuum at 10 inches Hg for 10 min.

# STEP 4: Dry the membranes of Filter Plate

- · Gently tap the tips of the Filter Plate on paper towel to remove residual liquid
- · Apply vacuum at 10 inches Hg for an addition 10 min.

# STEP 5: DNA Elution

- · place a 96-well PCR Plate (provided) on top of a clean 96-well plate (provided, the third 96-Well, 2 ml Plate). Place the combined plate on the rack of vacuum manifold and cover the manifold lid.
- $\cdot\,$  place the Filter Plate on top of the 96-well PCR Plate.
- $\cdot$  Add 50  $\sim$  100  $\mu$ l of Elution Buffer or ddH<sub>2</sub>O (pH8.0-8.5) to the membrane center of the Filter Plate. Stand for 3 min until Elution Buffer or ddH<sub>2</sub>O has been absorbed by the membrane completely.
- · Apply vacuum at 10 inches Hg for 5 min to elute DNA to the 96-well PCR plate.

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